



In vivo	PD-1-induced T-cell activation. [4]
Kinase Assay	The equilibrium binding experiments of fluorescent peptides to Bcl-xL protein were performed in an Analyst 96-well plate reader under the following conditions: each individual well in a 96-well assay plate contained 5 $\mu$ l DMSO, 15 nM fluorescent peptide, and increasing concentrations (from 0 to 2.24 $\mu$ M) of Bcl-xL protein in assay buffer in a final volume of 125 $\mu$ l. The plate was mixed on a shaker for 1 min and incubated at room temperature for an additional 15 min. The polarization in millipolarization units (mP) was measured at room temperature with an excitation wavelength at 485 nm and an emission wavelength at 530 nm. For assay stability testing, a plate containing a binding experiment was measured at different times over a 24-h period. Between each reading, the plate was covered with parafilm to prevent any solution evaporation. To determine the effect of DMSO on the assay, binding experiments were performed under conditions similar to those described above except that the amount of DMSO was varied from 0 to 4 to 8%. All experimental data were analyzed using Prism 3.0 software and Kd values were generated by fitting the experimental data using a sigmoidal dose-response nonlinear regression model [1].
Cell Research	RS4;11 cells were seeded at 50,000 per well in 96-well plates and treated with compounds diluted in half-log steps starting at 1 $\mu$ M and ending at 0.00005 $\mu$ M. All other leukemia and lymphoma cell lines were seeded at 15,000–20,000 cells per well in the appropriate medium and incubated with ABT-199 or navitoclax for 48 h. Effects on proliferation were determined using Cell TiterGlo reagent. EC50 values were determined by nonlinear regression analysis of the concentration-response data. Mouse FL5.12-BCL-2 and FL5.12-BCL-XL cells were propagated and assessed as described previously. Bak <sup>+/+</sup> Bax <sup>+/+</sup> double knockout mouse embryonic fibroblasts were seeded into 96-well microtiter plates at 5,000 cells per well in DMEM supplemented with 10% FBS. ABT-199 in the same culture medium was added in half-log dilutions starting at 5 $\mu$ M. The cells were then incubated at 37 °C (5% CO <sub>2</sub> ) for 48 h, and the effects on proliferation were determined using Cell TiterGlo reagent according to the manufacturer's instructions [1].
Animal Research	Female C.B-17 SCID mice (DoHH2 and Granta-519 xenografts) and female C.B-17 SCID-beige mice (RS4;11 and Toledo xenografts) were inoculated with 1 $\times$ 10 <sup>6</sup> (DoHH2) or 5 $\times$ 10 <sup>6</sup> (Granta-519, Toledo and RS4;11) cells subcutaneously in the right flank. The inoculation volume (0.2 ml) comprised a 50:50 mixture of cells in growth media and Matrigel. Electronic calipers were used to measure the length and width of each tumor 2–3 times per week. Tumor volume was estimated by applying the following equation: volume = length $\times$ width <sup>2</sup> /2. When tumors reached approximately 220 mm <sup>3</sup> , mice were size matched (day 0) into treatment and control groups. All xenograft trials were conducted using ten mice per group, and all mice were ear tagged and monitored individually throughout the studies. ABT-199 was formulated for oral dosing in 60% phosal 50 propylene glycol (PG), 30% polyethylene glycol (PEG) 400 and 10% ethanol, and bendamustine and rituximab were formulated in accordance with the manufacturer's instructions. ABT-199 was delivered approximately 2 h before bendamustine or bendamustine plus rituximab. TGImax was calculated as the greatest treatment response using the following equation: TGImax = (1 - mean tumor volume of the treated group/mean tumor volume of the vehicle control group) $\times$ 100. The TGD (%) was determined as the percentage increase of the median time period for the treatment group to reach an arbitrary tumor volume of 1,000 mm <sup>3</sup> relative to the vehicle control group. A complete tumor regression response was the portion of the population with tumors $\leq$ 25 mm <sup>3</sup> for at least three consecutive measurements [1].

## Solubility Information

Solubility	H2O: < 1 mg/mL (insoluble or slightly soluble), Ethanol: < 1 mg/mL (insoluble or slightly soluble), DMSO: 257.5 mg/mL (296.51 mM),Sonication is recommended. (< 1 mg/ml refers to the product slightly soluble or insoluble)
In vivo Formulation	10% DMSO+40% PEG300+5% Tween 80+45% Saline: 10 mg/mL (11.51 mM),Suspension. <i>Please add the solvents sequentially, clarifying the solution as much as possible before adding the next one. Dissolve by heating and/or sonication if necessary. Working solution is recommended to be prepared and used immediately. The formulation provided above is for reference purposes only. In vivo formulations may vary and should be modified based on specific experimental conditions.</i>

## Preparing Stock Solutions

	1mg	5mg	10mg
1 mM	1.1515 mL	5.7575 mL	11.5149 mL
5 mM	0.2303 mL	1.1515 mL	2.303 mL
10 mM	0.1151 mL	0.5757 mL	1.1515 mL
50 mM	0.023 mL	0.1151 mL	0.2303 mL

Please select the appropriate solvent to prepare the stock solution, according to the solubility of the product in different solvents. Please use it as soon as possible.

Note: The dilution table applies only to solid products. For liquid products, please calculate the stock solution based on the stated concentration and/or density.

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